

lence represents a local inflammatory reaction consisting of a high synovial white blood cell count.

Therefore, in the absence of an objective definition, it is difficult to consider purulence as a simple dichotomous variable. Subjective opinion of the surgeon regarding periprosthetic fluid can vary based on their clinical impression or concerns regarding the consequences of misdiagnosing PJIs. Moreover, PJI has a serious impact on patients' health and quality of life because patients may be subjected to additional surgical procedures and long-term antibiotic treatment. Therefore, surgeons should be cautious in applying subjective criteria for ruling in or ruling out PJIs in suspected patients.

## REFERENCES

- [1] Berbari EF, Hanssen AD, Duffy MC, Steckelberg JM, Ilstrup DM, Harmsen WS, et al. Risk factors for prosthetic joint infection: case-control study. *Clin Infect Dis*. 1998;27:1247-1254.
- [2] Parvizi J, Ghanem E, Menashe S, Barrack RL, Bauer TW. Periprosthetic infection: what are the diagnostic challenges? *J Bone Joint Surg Am*. 2006;88 Suppl 4:138-147. doi:10.2106/JBJS.F.00609.
- [3] Trampuz A, Piper KE, Jacobson MJ, Hanssen AD, Unni KK, Osmon DR, et al. Sonication of removed hip and knee prostheses for diagnosis of infection. *N Engl J Med*. 2007;357:654-663. doi:10.1056/NEJMoa061588.
- [4] Schinsky MF, Della Valle CJ, Sporer SM, Paprosky WG. Perioperative testing for joint infection in patients undergoing revision total hip arthroplasty. *J Bone Joint Surg Am*. 2008;90:1869-1875. doi:10.2106/JBJS.G.01255.
- [5] Osmon DR, Berbari EF, Berendt AR, Lew D, Zimmerli W, Steckelberg JM, et al. Diagnosis and management of prosthetic joint infection: clinical practice guidelines by the Infectious Diseases Society of America. *Clin Infect Dis*. 2013;56:e1-e25. doi:10.1093/cid/cis803.
- [6] Mangram AJ, Horan TC, Pearson ML, Silver LC, Jarvis WR. Guideline for prevention of surgical site infection, 1999. Centers for Disease Control and Prevention (CDC) Hospital Infection Control Practices Advisory Committee. *Am J Infect Control*. 1999;27:97-132; quiz 133-134; discussion 96.
- [7] Allami MK, Jamil W, Fourie B, Ashton V, Gregg PJ. Superficial incisional infection in arthroplasty of the lower limb. Interobserver reliability of the current diagnostic criteria. *J Bone Joint Surg Br*. 2005;87:1267-1271. doi:10.1302/0301-620X.87B9.16672.
- [8] Mikhael MM, Hanssen AD, Sierra RJ. Failure of metal-on-metal total hip arthroplasty mimicking hip infection. A report of two cases. *J Bone Joint Surg Am*. 2009;91:443-446. doi:10.2106/JBJS.H.00603.
- [9] Browne JA, Bechtold CD, Berry DJ, Hanssen AD, Lewallen DG. Failed metal-on-metal hip arthroplasties: a spectrum of clinical presentations and operative findings. *Clin Orthop Relat Res*. 2010;468:2313-2320. doi:10.1007/s11999-010-1419-0.
- [10] Engh CA, Ho H, Engh CA. Metal-on-metal hip arthroplasty: does early clinical outcome justify the chance of an adverse local tissue reaction? *Clin Orthop Relat Res*. 2010;468:406-412. doi:10.1007/s11999-009-1063-8.
- [11] Bonnaig NS, Freiberg RA, Freiberg AA. Total hip arthroplasty with ceramic-on-ceramic bearing failure from third-body wear. *Orthopedics*. 2011;34:132. doi:10.3928/01477447-20101221-36.
- [12] Kim TY, Kim SJ, Lee YK, Koo KH. Accumulation of fatty marrow in the osteonecrotic hip mimicking joint infection. *Clin Orthop Relat Res*. 2012;470:877-882. doi:10.1007/s11999-011-2048-y.
- [13] Malech HL, Deleo FR, Quinn MT. The role of neutrophils in the immune system: an overview. *Methods Mol Biol*. 2014;1124:3-10. doi:10.1007/978-1-62703-845-4\_1.
- [14] Dougherty SH. Pathobiology of infection in prosthetic devices. *Rev Infect Dis*. 1988;10:1102-1117.
- [15] Archibeck MJ, Rosenberg AG, Sheinkop MB, Berger RA, Jacobs JJ. Gout-induced arthropathy after total knee arthroplasty: a report of two cases. *Clin Orthop Relat Res*. 2001;377-382.
- [16] Jacobs JJ, Gilbert JL, Urban RM. Corrosion of metal orthopaedic implants. *J Bone Joint Surg Am*. 1998;80:268-282.
- [17] Judd KT, Noiseux N. Concomitant infection and local metal reaction in patients undergoing revision of metal on metal total hip arthroplasty. *Iowa Orthop J*. 2011;31:59-63.
- [18] Watters TS, Eward WC, Hallows RK, Dodd LG, Wellman SS, Bolognesi MP. Pseudotumor with superimposed periprosthetic infection following metal-on-metal total hip arthroplasty: a case report. *J Bone Joint Surg Am*. 2010;92:1666-1669. doi:10.2106/JBJS.I.01208.
- [19] Aljaniipour P, Adeli B, Hansen EN, Chen AF, Parvizi J. Intraoperative purulence is not reliable for diagnosing periprosthetic joint infection. *J Arthroplasty*. 2015;30:1403-1406. doi:10.1016/j.arth.2015.03.005.

● ● ● ● ●

**Authors:** Juan C. Martinez Pastor, Derek Amanatullah, Stuart Goodman, Ester Garcia Ultra, Marta Sabater Martos, Jake A. Mooney

## QUESTION 7: Is aseptic loosening (AL) associated with an undiagnosed periprosthetic joint infections (PJIs)?

**RECOMMENDATION:** Some percentage of AL is due to culture-negative infection, since up to 10% of culture-negative cases contain bacteria when screened by molecular methods. Whether this correlates to an undiagnosed infection causing AL remains unclear. Understanding this issue is limited by the ability of bacterial culture to function as an effective gold standard for detecting infection. The role of molecular techniques such as next generation sequencing in this setting needs to be explored.

**LEVEL OF EVIDENCE:** Limited

**DELEGATE VOTE:** Agree: 90%, Disagree: 8%, Abstain: 2% (Super Majority, Strong Consensus)

## RATIONALE

Loosening is one of the most common indications for total joint arthroplasty revision. Differentiating between PJI and AL is important in determining appropriate treatment. Loosening is considered aseptic when the radiographic or clinical findings associated with loosening are present in the absence of clinical or laboratory evidence of infection. Radiographic determination of loosening has an excellent specificity and positive predictive value, however, a poor sensitivity and negative predictive value, and thus should not be used to exclude loosening [1].

There is the possibility that microorganisms live on or around implants without signs or symptoms of infection, which can lead to AL. Several prospective and retrospective studies have supported that at least a fraction of cases with AL have been associated with

higher rates of bacterial growth. The reported prevalence of unexpected positive cultures (UPC) in presumed aseptic revision arthroplasty varies from 5.9 to 23.9% [2-14]. This major variation might be due to small sample size, different culturing protocols (detection of bacteriologic 16S ribosomal RNA by polymerase chain reaction, sonication fluid cultures and conventional techniques of fluid and soft tissue cultures), laboratory contamination rates, as well as the heterogeneity of patients included in each study (i.e., revisions for isolated polyethylene wear, dislocation, fracture and implant loosening) [2,5]. Kempthorne et al. reported a case-control prospective study comparing AL patients (cases) and patients undergoing revision surgery for other causes (control) with a positive culture rate of 15% [2].

Some authors have related early AL to hidden PJI [3,7,11]. Ribera et al. and Fernandez-Sampedro et al. have observed a correlation between microbiology and prosthesis-age, which supports the possibility of early loosening being caused by hidden PJIs [3,11]. Among the studies reported, there is no consensus about the prognostic impact of UPC. Some authors have shown that even a single positive intraoperative culture has been correlated to prosthetic joint failure, especially with early loosening [11,12]. On the other hand, Portillo et al. have found that the growth of low-virulence organisms in revisions for apparent AL is not associated with early prosthesis failure [8].

While traditional laboratory analysis to evaluate for infection consists of intraoperative culture of periprosthetic tissue or fluids, it has been well-established that microbial culture is an imperfect means of detecting bacteria, as culture has been shown to fail to detect bacteria in as many as 15% of clinically apparent infectious cases [15]. The increasing utilization of molecular methods in recent years has increased the incidence of bacterial detection in cases of AL. One study of 74 culture negative aseptic implants revealed the presence of bacteria in 9 (12%) after screening with polymerase chain reaction (PCR) assays [16].

The discrepancy between traditional culture methods and culture-independent molecular methods to detect bacterial infection in implants has been discussed extensively in the literature [17]. A number of proposed theories have been put forward to explain the absence of cultured bacteria in clinically infected cases, including the effects of prophylactic antibiotic treatment, growth behavior of biofilms and insufficient growth time to detect orthopaedic-specific pathogens. Regardless of the reason, detection via culture appears to be an inadequately sensitive diagnostic tool for periprosthetic joint infections.

A consistent limitation of studies that compare molecular techniques to culture is a failure to perform complete (deoxyribonucleic acid) DNA sequencing. Without this additional information, confirmation and agreement cannot be made between samples that are both culture and PCR-positive. Additionally, the etiology of culture negative and PCR-positive samples cannot be explored. Studies that have conducted full DNA sequencing have found significant discrepancies between the predominant species in culture versus those found via PCR analysis and the classic bacterial species that would be expected in PJIs [16]. The role of contamination in molecular methods also remains ill-defined. A carefully conducted study directly addressing this question found no significant difference in culture and 16S rRNA PCR of explanted implants [18].

An alternative theory to explain the phenomenon of culture-negative and PCR-positive clinically infected cases is the role of endotoxin. The detection limits for endotoxin are comparable to the stimulatory threshold, possibly resulting in unrecognized endotoxin [19]. Endotoxin alone replicates the effect of aseptic loosening [20] and can also adhere to titanium particles and implant surfaces [21]. In cases where bacteria are truly eradicated, cellular debris may create a false positive PCR, and residual endotoxin may initiate a local inflammatory response, resulting in culture negative loosening [22].

It is apparent that advanced modern molecular techniques detect bacteria in aseptic joints at a greater rate and with greater diversity than traditional microbial cultures. It is likely that a PJI is present in a greater number of cases with implant loosening than

previously suspected. More detailed studies are required to determine the true incidence of loosening due to infection and the exact pathogenic process that may differentiate culture and PCR-positive infections from culture-negative, but PCR-positive infections.

## REFERENCES

- Abrahams JM, Kim YS, Callary SA, et al. The diagnostic performance of radiographic criteria to detect aseptic acetabular component loosening after revision total hip arthroplasty. *Bone Joint J.* 2017;99B:458-464. doi:10.1302/0301-620X.99B4.BJ-2016-0804.R1.
- Kempthorne JT, Ailabouni R, Raniga S, Hammer D, Hooper G. Occult infection in aseptic joint loosening and the diagnostic role of implant sonication. *Biomed Res Int.* 2015;2015. doi:10.1155/2015/946215
- Ribera A, Morata L, Moranas J, et al. Clinical and microbiological findings in prosthetic joint replacement due to aseptic loosening. *J Infect.* 2014;69:235-243. doi:10.1016/j.jinf.2014.05.003.
- Padegimas EM, Lawrence C, Narzikul AC, et al. Future surgery after revision shoulder arthroplasty: the impact of unexpected positive cultures. *J Shoulder Elb Surg.* 2017;26:975-981. doi:10.1016/j.jse.2016.10.023.
- Barrack RL, Aggarwal A, Burnett RSJ, et al. The fate of the unexpected positive intraoperative cultures after revision total knee arthroplasty. *J Arthroplasty.* 2007;22:94-99. doi:10.1016/j.arth.2007.03.029.
- Jacobs AME, Bénard M, Meis JF, van Hellemond G, Goosen JHM. The unsuspected prosthetic joint infection. *Bone Joint J.* 2017;99-B:1482-1489. doi:10.1302/0301-620X.99B11.BJ-2016-0655.R2.
- Bereza PL, Ekiel A, Auguściak-Duma A, et al. Identification of asymptomatic prosthetic joint infection: microbiologic and operative treatment outcomes. *Surg Infect (Larchmt).* 2017;18:582-587. doi:10.1089/sur.2016.253.
- Portillo ME, Salvadó M, Alier A, et al. Prosthesis failure within 2 years of implantation is highly predictive of infection. *Clin Orthop Relat Res.* 2013;471:3672-3678. doi:10.1007/s11999-013-3200-7.
- Ince A, Rupp J, Frommelt L, Katzer A, Gille J, Löhr J. Is "aseptic" loosening of the prosthetic cup after total hip replacement due to nonculturable bacterial pathogens in patients with low-grade infection? *Clin Infect Dis.* 2004;39:1599-1603. doi:10.1086/425303.
- Staats K, Kolbitsch P, Sigmund IK, Hobusch GM, Holinka J, Windhager R. Outcome of total hip and total knee revision arthroplasty with minor infection criteria: a retrospective matched-pair analysis. *J Arthroplasty.* 2017;32:1266-1271. doi:10.1016/j.arth.2016.11.016.
- Fernandez-Sampedro M, Salas-Venero C, Fariñas-Álvarez C, et al. 26 postoperative diagnosis and outcome in patients with revision arthroplasty for aseptic loosening. *BMC Infect Dis.* 2015;15:232. doi:10.1186/s12879-015-0976-y.
- Saleh A, Guirguis A, Klika AK, Johnson L, Higuera CA, Barsoum WK. Unexpected positive intraoperative cultures in aseptic revision arthroplasty. *J Arthroplasty.* 2014;29:2181-2186. doi:10.1016/j.arth.2014.07.010
- Berend KR, Lombardi AVJ, Adams JB. Unexpected positive intraoperative cultures and gram stain in revision total hip arthroplasty for presumed aseptic failure. *Orthopedics.* 2007;30:1051-1053.
- Kelly JD, Hobgood ER. Positive culture rate in revision shoulder arthroplasty. *Clin Orthop Relat Res.* 2009;467:2343-2348. doi:10.1007/s11999-009-0875-x.
- Garvin K, Hanssen A. Infection after total hip arthroplasty. Past, present, and future. *J Bone Jt Surg Am.* 1995;77:1576-1588.
- Kobayashi N, Procop GW, Krebs V, Kobayashi H, Bauer TW. Molecular identification of bacteria from aseptically loose implants. *Clin Orthop Relat Res.* 2008;466:1716-1725. doi:10.1007/s11999-008-0263-y.
- Wasko MK, Goodman SB. Emperor's new clothes: is particle disease really infected particle disease? *J Orthop Res.* 2016;34:1497-1504. doi:10.1002/jor.23292
- Bjerkkan G, Witsø E, Nor A, et al. A comprehensive microbiological evaluation of fifty-four patients undergoing revision surgery due to prosthetic joint loosening. *J Med Microbiol.* 2012;61:572-581. doi:10.1099/jmm.0.036087-0
- Hitchins VM, Merritt K. Decontaminating particles exposed to bacterial endotoxin (LPS). *J Biomed Mater Res.* 1999;46:434-437. doi:10.1002/(SICI)1097-4636(19990905)46:3<434::AID-JBM17>3.0.CO;2-L.
- Bi Y, Seibold JM, Kaar SG, et al. Adherent endotoxin on orthopedic wear particles stimulates cytokine production and osteoclast differentiation. *J Bone Miner Res.* 2001;16:2082-2091. doi:10.1359/jbmr.2001.16.11.2082.
- Ragab AA, Van De Motter R, Lavish SA, et al. Measurement and removal of adherent endotoxin from titanium particles and implant surfaces. *J Orthop Res.* 1999;17:803-809. doi:10.1002/jor.1100170603.
- Sundfeldt M, Carlsson L V., Johansson CB, Thomsen P, Gretzer C. Aseptic loosening, not only a question of wear: a review of different theories. *Acta Orthop.* 2006;77:177-197. doi:10.1080/17453670610045902

