

Authors: Arnaldo Hernandez, Roberto Rossi

## QUESTION 1: Should culture samples be taken during arthroscopic treatment of a knee joint infection? If so, how many and from which area in the joint?

**RECOMMENDATION:** Yes, culture samples should be taken during arthroscopic treatment of a knee joint infection. We recommend that at least three culture samples from different sites be taken.

**LEVEL OF EVIDENCE:** Moderate

**DELEGATE VOTE:** Agree: 100%, Disagree: 0%, Abstain: 0% (Unanimous, Strongest Consensus)

### RATIONALE

Infections of the knee joint can occur either from hematogenous spread or directly due to local trauma or a medical intervention. Infections after an arthroscopy for anterior cruciate ligament reconstruction (ACLR) or meniscal pathology are reported throughout the literature [1–18]. Infection can also occur in healthy native knees [13,19–24]. Sending intraoperative samples of synovial fluid and tissue for microbiological analysis is commonly reported in the literature [1–12,14–24], with only one study reporting no intraoperative samples for culture [13]. Two studies described the number of samples taken during the arthroscopy [11,19]. In both of the studies, five samples were taken and sent for culture. Unfortunately, no studies described an optimal area of the joint from which to take the samples.

When considering the existing research, it can be concluded that samples should be taken during arthroscopic treatment for a knee infection. However, based on the review of the literature, no conclusion can be drawn about the number of samples.

There is more research describing the number of samples to be taken during debridement in periprosthetic joint infection (PJI). In their study on 113 PJIs, Gandhi et al. concluded that the optimal number of cultures needed to obtain a positive test result was four (specificity = 0.61, sensitivity = 0.63). Furthermore, they stated that increasing the number of samples increases specificity but reduces sensitivity [25]. In the same study, the samples were collected from representative areas of the joint, including, but not limited to, synovium, intramedullary tissue, prosthetic interface and tissue from the adjacent bone [25].

During the previous consensus meeting in 2013, it was concluded that three to six samples should be obtained intraoperatively in suspected PJI cases [26]. Similarly, other authors confirmed that three to five samples should be obtained from deep tissues during surgery for suspected PJI [27,28].

There is no agreement about the area of the joint the samples should be taken from during arthroscopic treatment of septic knee arthritis. In their review, Bauer et al. reported that the samples should be taken from the deep tissue [29]. In their systematic review, Mouzopoulos et al. suggested that during arthroscopic treatment of septic ACLR, samples for culture should be taken from multiple areas, such as synovial lining, graft, femoral and tibial tunnel [30].

Based on the available data, no definitive conclusion can be drawn on the number of samples needed and the area of the joint they should be taken from during arthroscopic treatment of septic knees. Studies based on PJI were considered, as well as literature reviewed on knee septic arthritis after ACLR. Based on this data, it may be extrapolated that at least three samples should be collected

during arthroscopic treatment of knee joint infection. Furthermore, they should be taken from multiple areas of the joint: graft, synovial lining and from the femoral and tibial tunnels when present. It is reasonable to also collect samples from other areas, such as the medial and lateral gutters and the suprapatellar pouch.

### REFERENCES

- [1] Abdel-Aziz A, Radwan YA, Rizk A. Multiple arthroscopic debridement and graft retention in septic knee arthritis after ACL reconstruction: a prospective case-control study. *Int Orthop*. 2014;38:73–82. doi:10.1007/s00264-013-2123-y.
- [2] D'Angelo GL, Ogilvie-Harris DJ. Septic arthritis following arthroscopy, with cost/benefit analysis of antibiotic prophylaxis. *Arthroscopy*. 1988;4:10–14.
- [3] Fong SY, Tan JL. Septic arthritis after arthroscopic anterior cruciate ligament reconstruction. *Ann Acad Med Singap*. 2004;33:228–234.
- [4] Hantes ME, Raoulis VA, Doxariotis N, Drakos A, Karachalios T, Malizos KN. Management of septic arthritis after arthroscopic anterior cruciate ligament reconstruction using a standard surgical protocol. *Knee*. 2017;24:588–593. doi:10.1016/j.knee.2017.02.007.
- [5] Indelli PF, Dillingham M, Fanton G, Schurman DJ. Septic arthritis in post-operative anterior cruciate ligament reconstruction. *Clin Orthop Relat Res*. 2002;182–188.
- [6] Judd D, Bottoni C, Kim D, Burke M, Hooker S. Infections following arthroscopic anterior cruciate ligament reconstruction. *Arthroscopy*. 2006;22:375–384. doi:10.1016/j.arthro.2005.12.002.
- [7] Kim HJ, Lee HJ, Lee JC, Min SG, Kyung HS. Evaluation of infection after anterior cruciate ligament reconstruction during a short period. *Knee Surg Relat Res*. 2017;29:45–51. doi:10.5792/ksrr.16.019.
- [8] Kubiak G, Fabiś J. Evaluation of treatment strategy of acute knee infection after ACL reconstruction with hamstring. *Pol Orthop Traumatol*. 2013;78:235–238.
- [9] McAllister DR, Parker RD, Cooper AE, Recht MP, Abate J. Outcomes of post-operative septic arthritis after anterior cruciate ligament reconstruction. *Am J Sports Med*. 1999;27:562–570. doi:10.1177/03635465990270050301.
- [10] Schollin-Borg M, Michaëlsson K, Rahme H. Presentation, outcome, and cause of septic arthritis after anterior cruciate ligament reconstruction: a case control study. *Arthroscopy*. 2003;19:941–947.
- [11] Schuster P, Schulz M, Immendoerfer M, Mayer P, Schlumberger M, Richter J. Septic arthritis after arthroscopic anterior cruciate ligament reconstruction: evaluation of an arthroscopic graft-retaining treatment protocol. *Am J Sports Med*. 2015;43:3005–3012. doi:10.1177/0363546515603054.
- [12] Sonnery-Cottet B, Archbold P, Zayni R, Bortolletto J, Thaanat M, Prost T, et al. Prevalence of septic arthritis after anterior cruciate ligament reconstruction among professional athletes. *Am J Sports Med*. 2011;39:2371–2376. doi:10.1177/0363546511417567.
- [13] Thiery JA. Arthroscopic drainage in septic arthritides of the knee: a multicenter study. *Arthroscopy*. 1989;5:65–69. doi:10.1016/0749-8063(89)90095-9.
- [14] Torres-Claramunt R, Pelfort X, Erquicia J, Gil-González S, Gelber PE, Puig L, et al. Knee joint infection after ACL reconstruction: prevalence, management and functional outcomes. *Knee Surg Sports Traumatol Arthrosc*. 2013;21:2844–2849. doi:10.1007/s00167-012-2264-3.
- [15] Van Tongel A, Stuyck J, Bellemans J, Vandenneucker H. Septic arthritis after arthroscopic anterior cruciate ligament reconstruction: a retrospective analysis of incidence, management and outcome. *American J Sports Med*. 2007;35:1059–1063. doi:10.1177/0363546507299443.
- [16] Wang C, Ao Y, Wang J, Hu Y, Cui G, Yu J. Septic arthritis after arthroscopic anterior cruciate ligament reconstruction: a retrospective analysis of inci-

- dence, presentation, treatment, and cause. *Arthroscopy*. 2009;25:243-249. doi:10.1016/j.arthro.2008.10.002.
- [17] Williams RJ, Laurencin CT, Warren RF, Speciale AC, Brause BD, O'Brien S. Septic arthritis after arthroscopic anterior cruciate ligament reconstruction. Diagnosis and management. *Am J Sports Med* 1997;25:261-7. doi:10.1177/036354659702500222.
- [18] Kuo C-L, Chang J-H, Wu C-C, Shen P-H, Wang C-C, Lin L-C, et al. Treatment of septic knee arthritis: comparison of arthroscopic debridement alone or combined with continuous closed irrigation-suction system. *J Trauma*. 2011;71:454-459. doi:10.1097/TA.0b013e3181ec4734.
- [19] Aim F, Delambre J, Bauer T, Hardy P. Efficacy of arthroscopic treatment for resolving infection in septic arthritis of native joints. *Orthop Traumatol Surg Res*. 2015;101:61-64. doi:10.1016/j.otsr.2014.11.010.
- [20] Balabaud L, Gaudias J, Boeri C, Jenny J-Y, Kehr P. Results of treatment of septic knee arthritis: a retrospective series of 40 cases. *Knee Surg Sports Traumatol Arthrosc*. 2007;15:387-392. doi:10.1007/s00167-006-0224-5.
- [21] Shukla A, Beniwal SK, Sinha S. Outcome of arthroscopic drainage and debridement with continuous suction irrigation technique in acute septic arthritis. *J Clin Orthop Trauma*. 2014;5:1-5. doi:10.1016/j.jcot.2014.01.004.
- [22] Stutz G, Kuster MS, Kleinstück F, Gächter A. Arthroscopic management of septic arthritis: stages of infection and results. *Knee Surg Sports Traumatol Arthrosc*. 2000;8:270-274.
- [23] Vispo Seara JL, Barthel T, Schmitz H, Eulert J. Arthroscopic treatment of septic joints: prognostic factors. *Arch Orthop Trauma Surg*. 2002;122:204-211. doi:10.1007/s00402-001-0386-z.
- [24] Yanmiş I, Ozkan H, Koca K, Kılıncıoğlu V, Bek D, Tunay S. The relation between the arthroscopic findings and functional outcomes in patients with septic arthritis of the knee joint, treated with arthroscopic debridement and irrigation. *Acta Orthop Traumatol Turc*. 2011;45:94-99. doi:10.3944/AOTT.2011.2258.
- [25] Gandhi R, Silverman E, Courtney PM, Lee GC. How many cultures are necessary to identify pathogens in the management of total hip and knee arthroplasty infections? *J Arthroplasty*. 2017;32:2825-2828. doi:10.1016/j.arth.2017.04.009.
- [26] Parvizi J, Gehrke T, Chen AF. Proceedings of the international consensus on periprosthetic joint infection. *Bone Joint J*. 2013;95-B:1450-1452. doi:10.1302/0301-620X.95B11.33135.
- [27] Parvizi J, Erkocak OF, Della Valle CJ. Culture-negative periprosthetic joint infection. *J Bone Joint Surg Am*. 2014;96:430-436. doi:10.2106/JBJS.L.01793.
- [28] Parvizi J, Adeli B, Zmistowski B, Restrepo C, Greenwald AS. Management of periprosthetic joint infection: the current knowledge: AAOS exhibit selection. *J Bone Joint Surg Am*. 2012;94:e104. doi:10.2106/JBJS.K.01417.
- [29] Bauer T, Boisrenoult P, Jenny JY. Post-arthroscopy septic arthritis: current data and practical recommendations. *Orthop Traumatol Surg Res*. 2015;101:S347-S350. doi:10.1016/j.otsr.2015.09.004.
- [30] Mouzopoulos G, Fotopoulos VC, Tzurbakis M. Septic knee arthritis following ACL reconstruction: a systematic review. *Knee Surg Sports Traumatol Arthrosc*. 2009;17:1033-1042. doi:10.1007/s00167-009-0793-1.

**Authors:** Sam Oussedik, Kevin Plancher, Ilaria Morelli, Domenico Ravier, Nimit Patel

## QUESTION 2: What diagnostic “algorithm” should be used to diagnose infection following anterior cruciate ligament reconstruction (ACLR)?

**RECOMMENDATION:** The “algorithm” to diagnose postoperative infection in patients with ACLR should include clinical presentation, serological tests including C-reactive protein (CRP) and erythrocyte sedimentation rate (ESR) and analysis of the synovial fluid aspirate including gram staining and culture.

**LEVEL OF EVIDENCE:** Moderate

**DELEGATE VOTE:** Agree: 100%, Disagree: 0%, Abstain: 0% (Unanimous, Strongest Consensus)

### RATIONALE

Postoperative infections following ACLR are rare, occurring in only 0.14–5.7% of cases [1–5]. As a result, clinical studies are limited and have small sample sizes. However, the general consensus is that the clinical presentation, laboratory blood tests, (specifically (CRP) and ESR) and synovial fluid aspiration analysis are essential for the diagnosis of infection after ACLR [6–13]. Magnetic resonance imaging can detect joint effusion, synovitis, edema of adjacent soft tissues and bone marrow, bone erosions, sinus tracts and soft tissue abscesses, though this has only been reported in one study [14].

Features of the clinical presentation that raise suspicion of infection include fever, malaise, sudden change in knee pain of moderate intensity, local incision drainage, local warmth, local swelling, erythema, decreased knee range of motion and inguinal lymph node enlargement, though each of these symptoms is not present in all cases [8,11,15–17].

Laboratory blood analysis should be included in the diagnosis of infection after ACLR. Interpretation of results can be challenging, as elevated levels are routinely seen postoperatively, (typically peaking by postoperative day three), as a result of the surgical trauma [3,7,13,18]. C-reactive protein levels, which increase within six to eight hours after infection, have been shown to have the highest sensitivity and specificity. Reported average C-reactive protein levels in patients after ACLR with knee infection range from 55.8 to 203 mg/L (range, 10–400 mg/L) (normal 0–0.5 mg/L) [11,15–17]. ESR levels typically rise within 24 to 48 hours [19–21]. Average ESR values in patients with knee infec-

tion after ACLR range from 57 to 76 mm/h (range, 9–108 mm/h) in the literature (normal 1–10 mm/h) [11,13,15,17,18]. Peripheral white blood cell count has also been shown to be elevated in patients with postoperative knee infection after ACLR (9.1 to 10.8 x 10<sup>9</sup>/L), though this is not a consistent finding in the majority of patients [13,15,17]. Polymorphonuclear neutrophils (average 71.7%) and fibrinogen levels (average 774.7 mg/mL) have also been assessed and shown to be elevated in patients with ACLR and postoperative knee infection [13].

Gross inspection of knee joint aspiration commonly reveals turbid, yellow-green synovial fluid.[3] Microbiological analysis of synovial fluid aspirate is the most widely studied diagnostic method for septic arthritis [1,6,8,9,19,22,23]. Analysis includes gram staining, leukocyte counts, aerobic and anaerobic cultures and antibiotic sensitivities [6,13]. Positive leukocyte counts of aspirated knee fluid in knee infections after ACLR have also been reported [average 91,000 (range 64,000 to 129,000)] [6,11]. Several retrospective studies have shown that in most cases synovial fluid bacterial cultures are positive to coagulase-negative Staphylococci (*Staphylococcus epidermidis*), *Staphylococcus aureus*, *Streptococcus non-hemolytic*, *Staphylococcus schleiferi*, *Escherichia coli* or *Propionibacterium* in acute septic arthrosis [6,11,13,15,17–19,23,24].

Overall, there is consensus that the diagnostic algorithm for postoperative knee infection following ACLR should include sudden change in history and presentation to include change in knee pain profile, swelling and range of motion, in addition to elevated CRP