

TABLE 1. Recommendations for treatment of resistant TB in pediatrics

	Initial Phase	Maintenance Phase
INH-mono-resistance TB	RIF + PZA + EMB (2 months)	RIF + PZA + EMB (4-7 months)
RIF-mono-resistance TB	INH + PZA + EMB + FQN (2 months)	INH + EMB + FQN (10-16 months)

INH, Isoniazid; EMB, Ethambutol; RIF, Rifampicin; PZA, Pyrazinamide; FQN, Fluoroquinolones; TB, Tuberculosis

the conventional four-drug regimen (Table 1) [3,7,8]. For multi-drug resistant (MDR) TB, all guidelines recommend a longer treatment period of up to 24 months with all four anti-TB drugs [3,7,9]. Evaluation of the organism's drug susceptibility profile should also be conducted [3,7,9].

While some authors have reported favorable results with chemotherapy and non-operative splinting of the affected joint(s), others have recommended debridement of focal bony involvement and arthroscopic or open synovectomy to decrease the overall bioburden of infected material [10,11].

Arthrodesis, especially of the hip joint, may be an option in the event of severe destruction of the joint secondary to infection [12]. Orthopaedic interventions in spinal TB may occasionally be recommended to prevent deformity of the spine in pediatric patients. These procedures may include surgical intervention, application of a brace or cast in addition to standard chemotherapy. Proper immobilization of the growing spine in pediatric patients may help achieve a solid fusion without surgical procedures.

Surgical intervention is reserved for patients with formation of a large anterior column abscess, severe kyphotic deformity or progressive spinal deformity despite chemotherapy [13,14].

REFERENCES

- [1] Schaberg T, Bauer T, Castell S, Dalhoff K, Detjen A, Diel R, et al. [Recommendations for therapy, chemoprevention and chemoprophylaxis of tuberculosis in adults and children. German Central Committee against Tuberculosis (DZK), German Respiratory Society (DGP)]. *Pneumol Stuttg Ger*. 2012;66:133-171. doi:10.1055/s-0031-1291619.
- [2] Rapid Advice: Treatment of Tuberculosis in Children. Geneva: World Health

- [3] Organization; 2010. Gale-Rowe M, Menzies D, Sutherland J, Wong T, editors, chapter authors. Highlights of the new 7th edition of the Canadian Tuberculosis Standards. *Can Commun Dis Rep*. 2014;40:113-116.
- [4] Guidance for National Tuberculosis Programmes on the management of tuberculosis in children. Chapter 1: introduction and diagnosis of tuberculosis in children. *Int J Tuberc Lung Dis*. 2006;10:1091-1097.
- [5] Safdar N, Hinderaker SG, Baloch NA, Enarson DA, Khan MA, Morkve O. Childhood tuberculosis deskguide and monitoring: an intervention to improve case management in Pakistan. *BMC Health Serv Res*. 2011;11:187. doi:10.1186/1472-6963-11-187.
- [6] Berti E, Galli L, Venturini E, de Martini M, Chiappini E. Tuberculosis in childhood: a systematic review of national and international guidelines. *BMC Infect Dis*. 2014;14:S3. doi:10.1186/1471-2334-14-S1-S3.
- [7] Red Book: 2012 Report of the Committee on Infectious Diseases. 29th Edition (2012). American Academy of Pediatrics. Elk Grove Village, IL. 2012.
- [8] Falzon D, Jaramillo E, Schünemann HJ, Arentz M, Bauer M, Bayona J, et al. WHO guidelines for the programmatic management of drug-resistant tuberculosis: 2011 update. *Eur Respir J*. 2011;38:516-528. doi:10.1183/09031936.00073611.
- [9] Voss LM, Australasian Subgroup in Paediatric Infectious Disease of the Australasian Society for Infectious Diseases; Australasian Paediatric Respiratory Group. Management of tuberculosis in children. *J Paediatr Child Health*. 2000;36:530-536.
- [10] Titov AG, Nakonechniy GD, Santavirta S, Serdobintzev MS, Mazurenko SI, Kontinen YT. Arthroscopic operations in joint tuberculosis. *The Knee*. 2004;11:57-62. doi:10.1016/S0968-0160(03)00035-8.
- [11] Agarwal A, Qureshi NA, Khan SA, Kumar P, Samaiya S. Tuberculosis of the foot and ankle in children. *J Orthop Surg Hong Kong*. 2011;19:213-217. doi:10.1177/230949901101900217.
- [12] OZdemir HM, Yensel U, Cevat Ogün T, Senaran H, Kutlu A. Arthrodesis for tuberculous coxarthrosis: good outcome in 32 adolescents. *Acta Orthop Scand*. 2004;75:430-433.
- [13] Jain AK. Tuberculosis of the spine: a fresh look at an old disease. *J Bone Joint Surg Br*. 2010;92:905-913. doi:10.1302/0301-620X.92B7.24668.
- [14] Zhang H-Q, Wang Y-X, Guo C-F, Liu J-Y, Wu J-H, Chen J, et al. One-stage posterior approach and combined interbody and posterior fusion for thoracolumbar spinal tuberculosis with kyphosis in children. *Orthopedics*. 2010;33:808. doi:10.3928/01477447-20100924-10.

Authors: Ali Parsa, Irene Kalbian

QUESTION 8: What is the role of host gene expression and severity of acute osteoarticular infection in children, especially methicillin-resistant *Staphylococcus aureus* (*S. aureus*), or MRSA, infection?

RECOMMENDATION: Unknown. The limited literature available suggests altered host gene transcription related to the balance of the body's adaptive and innate immune responses may increase pediatric patients' susceptibility to severe osteoarticular infection, particularly in cases of MRSA. However, much more investigation is needed to determine which genes are most useful and how they can be utilized to help physicians anticipate the course of infection in a given patient.

LEVEL OF EVIDENCE: Consensus

DELEGATE VOTE: Agree: 75%, Disagree: 3%, Abstain: 22% (Super Majority, Strong Consensus)

RATIONALE

The severity of osteoarticular infection in otherwise healthy children varies greatly, even in the setting of infection by the same pathogen. Some pediatric patients experience a mild course that allows them to be discharged after a few days of hospital admission with antibiotic therapy. Other patients experience a protracted course and require major surgical intervention as well as intensive care management [1–3]. The contribution of genetic mechanisms to this wide range of clinical manifestations has been investigated to a limited extent. A similar diversity in illness severity has been observed in neoplastic and rheumatologic disorders, where there is evidence that ribonucleic acid expression plays a role in the presentation of these conditions [4–6]. Chaussabel et al. used gene expression microarrays in patients with seven autoimmune-related conditions and identified transcriptional changes (“diagnostic signatures”) that could be used to distinguish between these respective conditions [7]. Identifying a parallel set of transcriptional diagnostic indicators for the severity of osteoarticular infection may enhance the ability of physicians to treat this condition.

S. aureus is one of the leading pathogens causing hospital-acquired infection and MRSA infection is associated with over 6,000 deaths/year in the United States [8]. In a series of 99 children hospitalized with *S. aureus* infection, investigators used microarray analysis to characterize the transcriptional profiles in whole blood. Significant heterogeneity was observed in host signatures and transcriptional changes were identified. Furthermore, this heterogeneity was found to be associated with a more severe course of disease. Overall, patients with invasive *S. aureus* infection had an exaggerated expression of genes associated with the innate immune response and a diminished expression of adaptive immunity [9].

Ardura et al. conducted a study comparing gene expression in peripheral blood monocyte cells (PBMC) between 53 children with invasive *S. aureus* infection and 24 healthy children. Analysis of PBMC gene expression showed that patients with invasive *S. aureus* had lower numbers of central memory CD4+ and CD8+ T-cells and increased numbers of CD14+ monocytes versus healthy controls [10]. Ramilo et al. compared the immune system response in patients with *Escherichia coli* infection versus those with *S. aureus* infection. Their findings support the specific pattern described by Ardura et al. They found that patients with *S. aureus* infection had altered host gene expression associated with their adaptive immune response [11]. Gaviria-Agudelo et al. reported on a cohort of 12 pediatric patients with acute hematogenous osteomyelitis caused by MRSA, and they identified specific genes which correlated with the severity of disease in the early hospitalization period. Among the five distinct genes that were identified, three were up-regulated (P2RX1, SORT1, RETN) and two were down-regulated (LOC641788, STAT 4). STAT4

down-regulation showed the strongest correlation with disease severity [12].

While these findings provide some initial evidence for the role of host gene expression in the severity of acute osteoarticular infection in children, the literature on this topic remains sparse. Further studies are needed to examine this connection, particularly studies with larger sample sizes. An enhanced understanding of host gene expression patterns and the transcriptome in osteoarticular infection could enable physicians to better anticipate the risk of developing chronic osteomyelitis and, ultimately, facilitate personalized patient management strategies.

REFERENCES

- [1] Holmig S, Copley L, Grande L, Wilson P. Deep venous thrombosis associated with osteomyelitis in children. *J Bone Joint Surg Am.* 2007;89:1517–1523.
- [2] Creel AM, Durham SH, Benner KW, Alten JA, Winkler M. Severe invasive community-associated methicillin-resistant *Staphylococcus aureus* infections in previously healthy children. *Pediatr Crit Care Med.* 2009;10:323–327. doi:10.1097/PCC.0b013e3181988798.
- [3] Gonzalez B, Martinez-Aguilar G, Hulten K, Hammerman W, Coss B, Avalos-Mishaan A, et al. Severe Staphylococcal sepsis in adolescents in the era of community-acquired methicillin-resistant *Staphylococcus aureus*. *Pediatrics.* 2005;115:642–648.
- [4] Garcia-Romo GS, Caielli S, Vega B, Connolly J, Allantaz F, Xu Z, et al. Netting neutrophils are major inducers of type I IFN production in pediatric systemic lupus erythematosus. *Sci Transl Med.* 2011;3:73ra20–73ra20. doi:10.1126/scitranslmed.3001201.
- [5] Chaussabel D, Quinn C, Shen J, Patel P, Glaser C, Baldwin N, et al. A modular analysis framework for blood genomics studies: application to systemic lupus erythematosus. *Immunity.* 2008;29:150–164. doi:10.1016/j.immuni.2008.05.012.
- [6] Golub T, Slonim D, Tamayo P, Huard C, Gaasenbeek M, Mesirov J, et al. Molecular classification of cancer: class discovery and class prediction by gene expression monitoring. *Science.* 1999;286:531–537. doi:10.1126/science.286.5439.531.
- [7] Chaussabel D. Analysis of significance patterns identifies ubiquitous and disease-specific gene-expression signatures in patient peripheral blood leukocytes. *Ann NY Acad Sci.* 2005;1062:146–154. doi:10.1196/annals.1358.017.
- [8] Klein E, Smith DL, Laxminarayan R. Hospitalizations and deaths caused by methicillin-resistant *Staphylococcus aureus*, United States, 1999–2005. *Emerg Infect Dis.* 2007;13:1840–1846. doi:10.3201/eid1312.070629.
- [9] Bancheureau R, Jordan-Villegas A, Ardura M, Mejias A, Baldwin N, Xu H, et al. Host immune transcriptional profiles reflect the variability in clinical disease manifestations in patients with *Staphylococcus aureus* infections. *PLoS ONE.* 2012;7:e34390. doi:10.1371/journal.pone.0034390.
- [10] Ardura MI, Bancheureau R, Mejias A, Di Pucchio T, Glaser C, Allantaz F, et al. Enhanced monocyte response and decreased central memory T cells in children with invasive *Staphylococcus aureus* infections. *PLoS ONE.* 2009;4:e5446. doi:10.1371/journal.pone.0005446.
- [11] Ramilo O, Allman W, Chung W, Mejias A, Ardura M, Glaser C, et al. Gene expression patterns in blood leukocytes discriminate patients with acute infections. *Blood.* 2007;109:2066–2077. doi:10.1182/blood-2006-02-002477.
- [12] Gaviria-Agudelo C, Carter K, Tareen N, Pascual V, Copley LA. Gene expression analysis of children with acute hematogenous osteomyelitis caused by methicillin-resistant *Staphylococcus aureus*: correlation with clinical severity of illness. *PLoS ONE.* 2014;9(7):e103523. doi:10.1371/journal.pone.0103523.

