

QUESTION 3: Do antibiotic coatings on implants reduce the rates of surgical site infections/periprosthetic joint infections (SSIs/PJIs)?

RECOMMENDATION: The use of antibacterial coatings on implants has been shown to reduce SSIs and/or PJIs based on in vitro and pre-clinical animal model studies. The use of antibiotic-coated implants in small series of patients appears to be encouraging. Larger-scale studies to prove the value of these technologies are needed.

LEVEL OF EVIDENCE: Limited

DELEGATE VOTE: Agree: 90%, Disagree: 6%, Abstain: 4% (Super Majority, Strong Consensus)

RATIONALE

Implanted biomaterials continue to play a key role in orthopaedic surgery. However, infections surrounding these implanted biomaterials remain a leading cause of failure, especially in total hip and knee arthroplasties [1–3]. The biofilm theory and its role in the propagation of bacterial growth is postulated to play a quintessential role in the etiology and pathogenesis of PJIs in modern-day total joint arthroplasties (TJAs) [4–8]. Surface roughness, hydrophobicity and electrostatic charge are important characteristics of implanted biomaterials that are exploited by bacteria to promote adherence [9,10]. Strategies proposed to reduce the rates of these complications have included the use of implants coated with antiseptic materials or antibiotic agents. Antibacterial coatings engineered for the surfaces of implanted biomaterials have been an evolving technology over the last three decades [11]. Romano et al. described ideal characteristics of future antibacterial coatings, namely that they would be proven in vivo by demonstrating acceptable antibacterial properties towards a large spectrum of organisms, easy handling, cost-effectiveness and lack of local or systemic toxicity while ensuring bone healing, on-growth or in-growth [9].

Antibacterial coatings can be categorized into three groups: (1) perioperative antibacterial local carriers or coatings (LCC), (2) passive surface finishing/modification (PSM) and (3) active surface finishing/modification (ASM) [9].

The first group, LCC, are antibacterial carriers or coatings that are applied to implants at the time of surgery. The most popular and well-studied vectors in this category include antibiotic-laden bone cement, used when coating intramedullary nails or total joint components [12]. Antibiotic-laden hydrogel that may be applied to the implant by the surgeon has been shown to reduce surgical site infections in a multicenter randomized controlled trial of 380 patients undergoing primary and revision total hip and total knee arthroplasties [13]. Similarly, a pilot study of second-stage implantation for prosthetic joint infections utilized implants coated with a resorbable calcium based bone substitute mixed with gentamycin or vancomycin [14]. At a minimum follow-up of one year, 95% of patients did not show any clinical signs of infections. However, no control group was used in this pilot study [14]. Furthermore, these studies, as well as other smaller cohorts that have been reported, are underpowered to make definitive recommendations for its widespread use.

The second group, PSM, revolves around the premise that chemical and/or physical modifications to the surface of an implanted biomaterial may reduce bacterial capabilities of adherence, and thus, prevent biofilm formations. These modifications are made without the planned release of bacteriostatic or bacteriocidal agents into the surrounding tissues. Such technology includes treatment of the surface layer of an implant with ultraviolet (UV) light irradiation to increase the hydrophilicity of the implant, which decreases bacterial adherence [15]. Changing the morphology of the surface layer of implants without decreasing the reliability of osseointegration has been proven capable of decreasing bacterial adherences in in vitro studies [16–19]. Polymer coatings (hydrophilic polymethacrylic acid or polyethylene oxide) or hydrogel coatings can also be applied to titanium implants, which helps deter bacterial adhesions [18,20–24]. PSM has great potential for future use on implanted biomaterials, however, there is concern regarding the osseointegration with coatings or surface modifications with strong anti-adhesive capabilities. Future in vitro and in vivo studies are needed prior to widespread clinical application.

The third group, ASM, includes modifications to the surface of the implant that impart pharmacologically-active antibacterial agents such as antibiotics, antiseptics, metal ions and/or organic compounds [9]. Antibacterial surface innovation largely revolves around metal ions such as magnesium, gold or silver [25–31], as well as non-metal elements such as chlorhexidine [32]. Antibiotics may be sprayed on or covalently bonded to the implant surface [33], applied via hydrogel or coating [13,34] or contained in and released via nanotubes [35,36]. While there is a myriad of vectors to deliver antibiotics to the surrounding tissue, there is a paucity of conclusive in vitro studies, and a relative lack of in vivo studies demonstrating safety and efficacy with this technology. Further confounding ASM is the wide variability of coatings studied. This makes it tremendously difficult to draw conclusions from the current literature regarding ASM. While studies have shown that antibiotic coatings do not affect bone healing in animal models [37,38], this technology has not been studied clinically.

Perhaps the most well-studied antibacterial coating are antiseptics, such as metal ions impregnated into the implant or applied via coating. Both in vitro and in vivo animal models have demonstrated significant antibacterial effects [23,25,26,28,31,36,39–41]. Additionally, clinical studies of silver-coated endoprostheses have demonstrated the efficacious antiseptic effects of the metal-ion coating in reducing infection [42–44]. However, these studies are largely retrospective in nature, and underpowered to render conclusive evidence supporting the widespread application of such technologies. While there are concerns of metal-ion toxicity that may result from such coatings, several studies have demonstrated little to no evidence of toxicity or side-effects [30,40,45]. Metal-ion coatings appear to be the most promising in terms of efficacy and near-future implementation based on review of the present literature surrounding antibacterial coatings.

Despite the promise of these individual reports, the paucity of high-level controlled trials in the setting of arthroplasty, suggests that it is too early to conclude that antibiotic coatings will reduce the rates of SSIs/PJIs following primary or revision procedures. However, these strategies could prove to be beneficial in high-risk primary or revision cases. Further high-quality studies are needed to address these questions.

REFERENCES

- [1] Melvin JS, Karthikeyan T, Cope R, Fehring TK. Early failures in total hip arthroplasty — a changing paradigm. *J Arthroplasty*. 2014;29:1285–1288. doi:10.1016/j.arth.2013.12.024.
- [2] Sharkey PF, Lichstein PM, Shen C, Tokarski AT, Parvizi J. Why are total knee arthroplasties failing today—has anything changed after 10 years? *J Arthroplasty*. 2014;29:1774–1778. doi:10.1016/j.arth.2013.07.024.
- [3] Khan M, Osman K, Green G, Haddad FS. The epidemiology of failure in total knee arthroplasty: avoiding your next revision. *Bone Joint J*. 2016;98–B:105–112. doi:10.1302/0301–620X.98B1.36293.
- [4] Waldvogel FA, Medoff G, Swartz MN. Osteomyelitis: a review of clinical features, therapeutic considerations and unusual aspects. *N Engl J Med*. 1970;282:198–206. doi:10.1056/NEJM197001222820406.
- [5] Griffin JW, Guillot SJ, Redick JA, Browne JA. Removed antibiotic-impregnated cement spacers in two-stage revision joint arthroplasty do not show biofilm formation in vivo. *J Arthroplasty*. 2012;27:1796–1799. doi:10.1016/j.arth.2012.06.019.
- [6] Nguyen LL, Nelson CL, Saccente M, Smeltzer MS, Wassell DL, McLaren SG. Detecting bacterial colonization of implanted orthopaedic devices by ultrasonication. *Clin Orthop Relat Res*. 2002;29–37.
- [7] Stoodley P, Nistico L, Johnson S, Lasko L-A, Baratz M, Gahlot V, et al. Direct demonstration of viable staphylococcus aureus biofilms in an infected total joint arthroplasty. A case report. *J Bone Joint Surg Am*. 2008;90:1751–1758. doi:10.2106/JBJS.G.00838.
- [8] Urish KL, DeMuth PW, Kwan BW, Craft DW, Ma D, Haider H, et al. Antibiotic-tolerant staphylococcus aureus biofilm persists on arthroplasty materials. *Clin Orthop Relat Res*. 2016;474:1649–1656. doi:10.1007/s11999–016–4720–8.
- [9] Romanò CL, Scarponi S, Gallazzi E, Romanò D, Drago L. Antibacterial coating of implants in orthopaedics and trauma: a classification proposal in an evolving panorama. *J Orthop Surg*. 2015;10:157. doi:10.1186/s13018–015–0294–5.
- [10] Chen Y, Busscher HJ, van der Mei HC, Norde W. Statistical analysis of long- and short-range forces involved in bacterial adhesion to substratum surfaces as measured using atomic force microscopy. *Appl Environ Microbiol*. 2011;77:5065–5070. doi:10.1128/AEM.00502–11.
- [11] Gristina AG, Naylor P, Myrvik Q. Infections from biomaterials and implants: a race for the surface. *Med Prog Technol*. 1988;14:205–224.
- [12] Schmidmaier G, Kerstan M, Schwabe P, Südkamp N, Raschke M. Clinical experiences in the use of a gentamicin-coated titanium nail in tibia fractures. *Injury*. 2017;48:2235–2241. doi:10.1016/j.injury.2017.07.008.
- [13] Romanò CL, Malizos K, Capuano N, Mezzoprete R, D'Arienzo M, et al. Does an antibiotic-loaded hydrogel coating reduce early post-surgical infection after joint arthroplasty? *J Bone Joint Infect*. 2016;1:34–41. doi:10.7150/jbji.15986.
- [14] Logoluso N, Drago L, Gallazzi E, George DA, Morelli I, Romanò CL. Calcium-based, antibiotic-loaded bone substitute as an implant coating: a pilot clinical study. *J Bone Jt Infect*. 2016;1:59–64. doi:10.7150/jbji.17586.
- [15] Gallardo-Moreno AM, Pacha-Olivenza MA, Saldaña L, Pérez-Giraldo C, Bruque JM, Vilaboa N, et al. In vitro biocompatibility and bacterial adhesion of physico-chemically modified Ti6Al4V surface by means of UV irradiation. *Acta Biomater*. 2009;5:181–192. doi:10.1016/j.actbio.2008.07.028.
- [16] Della Valle C, Visai L, Santin M, Cigada A, Candiani G, Pezzoli D, et al. A novel antibacterial modification treatment of titanium capable to improve osseointegration. *Int J Artif Organs*. 2012;35:864–875. doi:10.5301/ijao.5000161.
- [17] Liu L, Bhatia R, Webster TJ. Atomic layer deposition of nano-TiO₂ thin films with enhanced biocompatibility and antimicrobial activity for orthopedic implants. *Int J Nanomedicine*. 2017;12:8711–8723. doi:10.2147/IJN.S148065.
- [18] Ma Y, Chen M, Jones JE, Ritts AC, Yu Q, Sun H. Inhibition of Staphylococcus epidermidis biofilm by trimethylsilane plasma coating. *Antimicrob Agents Chemother*. 2012;56:5923–5937. doi:10.1128/AAC.01739–12.
- [19] Diefenbeck M, Mückley T, Schrader C, Schmidt J, Zankovych S, Bossert J, et al. The effect of plasma chemical oxidation of titanium alloy on bone-implant contact in rats. *Biomaterials*. 2011;32:8041–8047. doi:10.1016/j.biomaterials.2011.07.046.
- [20] Drago L, Boot W, Dimas K, Malizos K, Hänsch GM, Stuyck J, et al. Does implant coating with antibacterial-loaded hydrogel reduce bacterial colonization and biofilm formation in vitro? *Clin Orthop Relat Res*. 2014;472:3311–3323. doi:10.1007/s11999–014–3558–1.
- [21] Pfeufer NY, Hofmann-Peiker K, Mühle M, Warnke PH, Weigel MC, Kleine M. Bioactive coating of titanium surfaces with recombinant human β -defensin-2 (rHu β D2) may prevent bacterial colonization in orthopaedic surgery. *J Bone Joint Surg Am*. 2011;93:840–846. doi:10.2106/JBJS.I.01738.
- [22] Chen R, Willcox MD, Ho KK, Smyth D, Kumar N. Antimicrobial peptide melimine coating for titanium and its in vivo antibacterial activity in rodent subcutaneous infection models. *Biomaterials*. 2016;85:142–151. doi:10.1016/j.biomaterials.2016.01.063.
- [23] Harris LG, Tosatti S, Wieland M, Textor M, Richards RG. Staphylococcus aureus adhesion to titanium oxide surfaces coated with non-functionalized and peptide-functionalized poly(L-lysine)-grafted-poly(ethylene glycol) copolymers. *Biomaterials*. 2004;25:4135–4148. doi:10.1016/j.biomaterials.2003.11.033.
- [24] Kazemzadeh-Narbat M, Noordin S, Masri BA, Garbuz DS, Duncan CP, et al. Drug release and bone growth studies of antimicrobial peptide-loaded calcium phosphate coating on titanium. *J Biomed Mater Res B Appl Biomater*. 2012;100:1344–1352. doi:10.1002/jbm.b.32701.
- [25] Kose N, Otuzbir A, Pekşen C, Kiremitçi A, Doğan A. A silver ion-doped calcium phosphate-based ceramic nanopowder-coated prosthesis increased infection resistance. *Clin Orthop Relat Res*. 2013;471:2532–2539. doi:10.1007/s11999–013–2894–x.
- [26] Kuehl R, Brunetto PS, Woischnig A-K, Varisco M, Rajacic Z, Vosbeck J, et al. Preventing implant-associated infections by silver coating. *Antimicrob Agents Chemother*. 2016;60:2467–2475. doi:10.1128/AAC.02934–15.
- [27] Mauerer A, Lange B, Welsch GH, Heidenau F, Adler W, Forst R, et al. Release of Cu²⁺ from a copper-filled TiO₂ coating in a rabbit model for total knee arthroplasty. *J Mater Sci Mater Med*. 2014;25:813–821. doi:10.1007/s10856–013–5116–x.
- [28] Norambuena GA, Patel R, Karau M, Wyles CC, Jannetto PJ, Bennet KE, et al. Antibacterial and biocompatible titanium-copper oxide coating may be a potential strategy to reduce periprosthetic infection: an in vitro study. *Clin Orthop Relat Res*. 2017;475:722–732. doi:10.1007/s11999–016–4713–7.
- [29] Shimazaki T, Miyamoto H, Ando Y, Noda I, Yonekura Y, Kawano S, et al. In vivo antibacterial and silver-releasing properties of novel thermal sprayed silver-containing hydroxyapatite coating. *J Biomed Mater Res B Appl Biomater*. 2010;92:386–389. doi:10.1002/jbm.b.31526.
- [30] Tsukamoto M, Miyamoto H, Ando Y, Noda I, Eto S, Akiyama T, et al. Acute and subacute toxicity in vivo of thermal-sprayed silver containing hydroxyapatite coating in rat tibia. *BioMed Res Int*. 2014;2014:902343. doi:10.1155/2014/902343.
- [31] Tran N, Kelley MN, Tran PA, Garcia DR, Jarrell JD, Hayda RA, et al. Silver doped titanium oxide-PDMS hybrid coating inhibits Staphylococcus aureus and Staphylococcus epidermidis growth on PEEK. *Mater Sci Eng C Mater Biol Appl*. 2015;49:201–209. doi:10.1016/j.msec.2014.12.072.
- [32] Riool M, Dirks AJ, Jaspers V, de Boer L, Loontjens TJ, van der Loos CM, et al. A chlorhexidine-releasing epoxy-based coating on titanium implants prevents Staphylococcus aureus experimental biomaterial-associated infection. *Eur Cell Mater*. 2017;33:143–157. doi:10.22203/eCM.v033a11.
- [33] Gerits E, Kucharíková S, Van Dijk P, Erdtmann M, Krona A, Lövenklev M, et al. Antibacterial activity of a new broad-spectrum antibiotic covalently bound to titanium surfaces. *J Orthop Res*. 2016;34:2191–2198. doi:10.1002/jor.23238.
- [34] Malizos K, Blauth M, Danita A, Capuano N, Mezzoprete R, Logoluso N, et al. Fast-resorbable antibiotic-loaded hydrogel coating to reduce post-surgical infection after internal osteosynthesis: a multicenter randomized controlled trial. *J Orthop Traumatol*. 2017;18:159–169. doi:10.1007/s10195–017–0442–2.
- [35] Ambrose CG, Clyburn TA, Mika J, Gogola GR, Kaplan HB, Wanger A, et al. Evaluation of antibiotic-impregnated microspheres for the prevention of implant-associated orthopaedic infections. *J Bone Joint Surg Am*. 2014;96:128–134. doi:10.2106/JBJS.L.01750.
- [36] Esfandiari N, Simchi A, Bagheri R. Size tuning of Ag-decorated TiO₂ nanotube arrays for improved bactericidal capacity of orthopedic implants. *J Biomed Mater Res A*. 2014;102:2625–2635. doi:10.1002/jbm.a.34934.
- [37] Moojen DJF, Vogely HC, Fleer A, Nikkels PGJ, Higham PA, Verbout AJ, et al. Prophylaxis of infection and effects on osseointegration using a tobramycin-periapatite coating on titanium implants—an experimental study in the rabbit. *J Orthop Res*. 2009;27:710–716. doi:10.1002/jor.20808.
- [38] Fassbender M, Minkwitz S, Kronbach Z, Strobel C, Kadow-Romacker A, Schmidmaier G, et al. Local gentamicin application does not interfere with bone healing in a rat model. *Bone*. 2013;55:298–304. doi:10.1016/j.bone.2013.04.018.

- [39] Cheng H, Li Y, Huo K, Gao B, Xiong W. Long-lasting in vivo and in vitro antibacterial ability of nanostructured titania coating incorporated with silver nanoparticles. *J Biomed Mater Res A*. 2014;102:3488–3499. doi:10.1002/jbm.a.35019.
- [40] Gosheger G, Hantes J, Ahrens H, Streitburger A, Buerger H, Erren M, et al. Silver-coated megaendoprostheses in a rabbit model—an analysis of the infection rate and toxicological side effects. *Biomaterials*. 2004;25:5547–5556. doi:10.1016/j.biomaterials.2004.01.008.
- [41] Kose N, Çaylak R, Pekşen C, Kiremitçi A, Burukoglu D, Koparal S, et al. Silver ion doped ceramic nano-powder coated nails prevent infection in open fractures: In vivo study. *Injury*. 2016;47:320–324. doi:10.1016/j.injury.2015.10.006.
- [42] Donati F, Di Giacomo G, D’Adamio S, Ziranu A, Careri S, Rosa M, et al. Silver-coated hip megaprosthesis in oncological limb salvage surgery. *BioMed Res Int*. 2016;2016:9079041. doi:10.1155/2016/9079041.
- [43] Hantes J, von Eiff C, Streitburger A, Balke M, Budny T, Henrichs MP, et al. Reduction of periprosthetic infection with silver-coated megaprostheses in patients with bone sarcoma. *J Surg Oncol*. 2010;101:389–395. doi:10.1002/jso.21498.
- [44] Wafa H, Grimer RJ, Reddy K, Jeys L, Abudu A, Carter SR, et al. Retrospective evaluation of the incidence of early periprosthetic infection with silver-treated endoprostheses in high-risk patients: case-control study. *Bone Joint J*. 2015;97-B:252–257. doi:10.1302/0301-620X.97B2.34554.
- [45] Scocianti G, Frenos F, Beltrami G, Campanacci DA, Capanna R. Levels of silver ions in body fluids and clinical results in silver-coated megaprostheses after tumour, trauma or failed arthroplasty. *Injury*. 2016;47 Suppl 4:S11–S16. doi:10.1016/j.injury.2016.07.042.

